UCMR4 Sampling Instructions
Anatek Labs, Inc.
www.anateklabs.com
208-883-2839 (Moscow) – 509-838-3999 (Spokane)

Contact us with any questions before sampling. Samples will be rejected if improperly submitted.

Clearly label each sample container. Use a permanent marker (Sharpie or similar) to indicate the sampling location and the date and time collected.

Use the proper container to collect your sample and do not rinse out any preservatives present. Specific sample containers are required for all UCMR4 analyses. Samples will be rejected if collected in the incorrect container and will require re-sampling. If you have any questions, please contact us before sampling (**1-208-883-2839**).

Refrigerate samples or place samples on ice immediately after sampling prior to shipping to lab. Samples must be submitted to the lab within 48 hours of sampling. UCMR4 protocols have strict preservation and temperature requirements thus samples must arrive at the laboratory cold (<10^{o}C) and in the correct containers or samples will be rejected.

Before submitting your samples to the lab, complete a chain of custody form for each point (EP), distribution system (DS), and water source (SR) . The additional 'two bottles per water system' noted on some tests are collected to allow the laboratory to run matrix spike samples required by UCMR4.

EPA 200.8 Metals in Drinking Water

Bottles: 250 mL HDPE plastic (one bottle for each Entry Point)

Fill the bottle under a slow stream of water until the bottle is filled up to the neck.

EPA 525.3 Pesticides

Bottles: 1 L amber glass (three bottles for each Entry Point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 530 Semivolatiles

Bottles: 1L amber glass (two bottles for each Entry Point, plus an additional two bottles per water system)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 541 Alcohols

Bottles: 60 mL amber glass vials (two per each Entry Point, plus an additional two bottles per water system)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 552.3: Haloacetic Acids

Bottles: 60 mL amber glass vials (two for each Distribution System, plus an additional two bottles per water system)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

SM 5310C: Total Organic Carbon (TOC)

Bottles: 2 x 40 mL amber glass vials (two for each Water Source)

EPA 300.0: Bromide

Bottles: 125 mL HDPE plastic (one for each Water Source)

EPA 544: Microcystins & Nodularin

Bottles: 550 mL amber glass (three bottles for each Entry Point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 545: Cylindrospermopsin & Anatoxin-a

Bottles: 125 mL amber glass (one bottle for each Entry Point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.

EPA 546 (ELISA): Total Microcystins

Bottles: 125 mL amber glass (one bottle for each Entry Point)

Fill the bottle under a slow stream of water until it is filled up to the neck. Gently replace the cap, tighten securely, and invert the sample a few times to dissolve the preservative.